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EXAMINER

SALMON, KATHERINE D

ART UNIT

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1634

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**Please find below and/or attached an Office communication concerning this application or proceeding.**

The time period for reply, if any, is set in the attached communication.

<b>Office Action Summary</b>	<b>Application No.</b> 10/748,374	<b>Applicant(s)</b> SU, XING	
	<b>Examiner</b> KATHERINE SALMON	<b>Art Unit</b> 1634	

-- The MAILING DATE of this communication appears on the cover sheet with the correspondence address --

### Period for Reply

A SHORTENED STATUTORY PERIOD FOR REPLY IS SET TO EXPIRE 3 MONTH(S) OR THIRTY (30) DAYS, WHICHEVER IS LONGER, FROM THE MAILING DATE OF THIS COMMUNICATION.

- Extensions of time may be available under the provisions of 37 CFR 1.136(a). In no event, however, may a reply be timely filed after SIX (6) MONTHS from the mailing date of this communication.
- If NO period for reply is specified above, the maximum statutory period will apply and will expire SIX (6) MONTHS from the mailing date of this communication.
- Failure to reply within the set or extended period for reply will, by statute, cause the application to become ABANDONED (35 U.S.C. § 133). Any reply received by the Office later than three months after the mailing date of this communication, even if timely filed, may reduce any earned patent term adjustment. See 37 CFR 1.704(b).

### Status

- 1) ☒ Responsive to communication(s) filed on 14 August 2008.
- 2a) ☐ This action is **FINAL**.                      2b) ☒ This action is non-final.
- 3) ☐ Since this application is in condition for allowance except for formal matters, prosecution as to the merits is closed in accordance with the practice under *Ex parte Quayle*, 1935 C.D. 11, 453 O.G. 213.

### Disposition of Claims

- 4) ☒ Claim(s) 1-17, 22-34, 36-38 and 41-45 is/are pending in the application.
- 4a) Of the above claim(s) \_\_\_\_\_ is/are withdrawn from consideration.
- 5) ☐ Claim(s) \_\_\_\_\_ is/are allowed.
- 6) ☒ Claim(s) 1-17, 22-34, 36-38 and 41-45 is/are rejected.
- 7) ☐ Claim(s) \_\_\_\_\_ is/are objected to.
- 8) ☐ Claim(s) \_\_\_\_\_ are subject to restriction and/or election requirement.

### Application Papers

- 9) ☐ The specification is objected to by the Examiner.
- 10) ☐ The drawing(s) filed on \_\_\_\_\_ is/are: a) ☐ accepted or b) ☐ objected to by the Examiner.  
Applicant may not request that any objection to the drawing(s) be held in abeyance. See 37 CFR 1.85(a).  
Replacement drawing sheet(s) including the correction is required if the drawing(s) is objected to. See 37 CFR 1.121(d).
- 11) ☐ The oath or declaration is objected to by the Examiner. Note the attached Office Action or form PTO-152.

### Priority under 35 U.S.C. § 119

- 12) ☐ Acknowledgment is made of a claim for foreign priority under 35 U.S.C. § 119(a)-(d) or (f).
- a) ☐ All    b) ☐ Some \*    c) ☐ None of:
- ☐ Certified copies of the priority documents have been received.
  - ☐ Certified copies of the priority documents have been received in Application No. \_\_\_\_\_.
  - ☐ Copies of the certified copies of the priority documents have been received in this National Stage application from the International Bureau (PCT Rule 17.2(a)).

\* See the attached detailed Office action for a list of the certified copies not received.

### Attachment(s)

- |  |   |
|--|---|
| 1) <input checked="" type="checkbox"/> Notice of References Cited (PTO-892)          | 4) <input type="checkbox"/> Interview Summary (PTO-413)           |
| 2) <input type="checkbox"/> Notice of Draftsperson's Patent Drawing Review (PTO-948) | Paper No(s)/Mail Date. _____                                      |
| 3) <input type="checkbox"/> Information Disclosure Statement(s) (PTO/SB/08)          | 5) <input type="checkbox"/> Notice of Informal Patent Application |
| Paper No(s)/Mail Date _____  | 6) <input type="checkbox"/> Other: _____                          |

### **DETAILED ACTION**

1. This action is in response to papers filed 8/14/2008.
2. Currently claims 1-17, 22-34, 36-38, 41-45 are pending. Claims 18-21, 35, and 39-40 have been cancelled.
3. The following rejections are newly applied as such the following rejection is Nonfinal.

### **Withdrawn Rejections**

4. The rejection of the claims under 35 USC 112/New Matter made in section 6 of the previous office action is moot based on amendments to the claims.
5. The rejection of claims 39-40 under 35 USC 112/second paragraph made in section 7 of the previous office action is moot based on the cancellation of the claims.
6. The rejection of the claims under the art as set forth in the 35 USC 103(a) made in sections 10-25 has been withdrawn. It is noted that the art of record has been used in the rejections below, but based upon new limitations and cited passages not made previously of record.

### **Claim Rejections - 35 USC § 112**

The following is a quotation of the second paragraph of 35 U.S.C. 112:

The specification shall conclude with one or more claims particularly pointing out and distinctly claiming the subject matter which the applicant regards as his invention.

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4. Claim 34 is rejected under 35 U.S.C. 112, second paragraph, as being indefinite for failing to particularly point out and distinctly claim the subject matter which applicant regards as the invention.

Claim 34 is unclear over the phrase “comprises less than 5 purine residues positively charged enhancer is an amine group”. It is not clear if the nucleic acid comprises less than 5 purine residues or if the positively charged enhance comprises less than 5 purine residues.

### ***Claim Rejections - 35 USC § 102***

The following is a quotation of the appropriate paragraphs of 35 U.S.C. 102 that form the basis for the rejections under this section made in this Office action:

A person shall be entitled to a patent unless –

(e) the invention was described in (1) an application for patent, published under section 122(b), by another filed in the United States before the invention by the applicant for patent or (2) a patent granted on an application for patent by another filed in the United States before the invention by the applicant for patent, except that an international application filed under the treaty defined in section 351(a) shall have the effects for purposes of this subsection of an application filed in the United States only if the international application designated the United States and was published under Article 21(2) of such treaty in the English language.

5. Claims 1-2, 5-7, 9-10, 13-17, 37-38, 42, 44-45 are rejected under 35

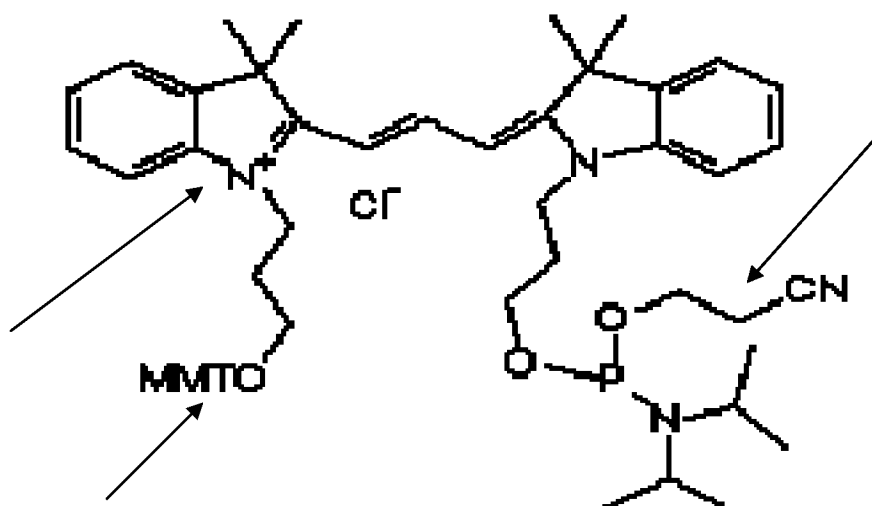
U.S.C. 102(e) as being anticipated by Mirkin et al. (US Patent Application Publication 2003/0211488 A1 November 13, 2003) as evidenced by Glen Research Catalog (Catalog Number 105913 [www.glenresearch.com](http://www.glenresearch.com)).

With regard to Claim 1, Mirkin et al. teaches a multiplexed detection method of oligonucleotide targets bound to capture probes and detected using SERS (surface enhanced Raman) Raman probes (Abstract, p. 3 paragraphs 45-49). Mirkin et al. teaches contacting a target nucleic acid with a plurality of capture probes bound to a substrate forming a single-stranded overhang (Abstract, p. 3 paragraphs 45-49). Mirkin et al. teaches contacting the probe-target with a population of Raman-active oligonucleotides which forms a three-component sandwich assay used in a microarray (e.g. biochip) format composed of nanoparticle probes (Raman probes) detecting a bound target: capture probe duplex (Abstract, p. 3 paragraphs 45-49 ).

Mirkin et al. teaches a method in which the probe has a nanoparticle- Cy3-labeled alkylthiol capped oligonucleotide (Example 2, p. 10). The amine group of the Cy3 label is considered a positively charged Raman signal enhancer because it maintains a positive charge on the amine group even after attachment of the complementary target.

Mirkin et al. teaches the method of attaching the Cy3 linker to the complex (examples 2 p. 10). Mirkin et al teaches the Cy3 modified label was purchased from Glen Research (paragraph 158 p. 10).

The following Cy3 chemical structure is from the Glen Research Catalog.



Mirkin et al. teaches that the oligonucleotide is attached to the phosphoramidite and the linker is attached to the DMT (displayed as MMTO on the figure) (p. 10 paragraph 158). Therefore the positively charged amine is not affected by the attachment of the linker and the oligonucleotide and would maintain its positive charge after binding to with the probe-target complex. The amine group on the Cy3 label is one “positively charged Raman signal enhancer” comprised in the Raman active probes.

Further Mirkin et al. teaches that nanoparticles can be made of silver or TiO<sub>2</sub> (p. 7 paragraph 116) which are both positively charged. The instant specification asserts that a positively charge enhancer can be any positively charged group that is attached to the oligonucleotides without blocking the binding of the oligonucleotide to a complementary sequence (paragraph 24 p. 6). The nanoparticle does not block the binding of the probe and the complementary sequence, has a positive charge and therefore is also a positively charged enhancer.

Therefore either the positively charged amine of Cy3 would be considered the positively charged Raman signal enhancer or the use of silver or TiO<sub>2</sub> would encompass the positively charged Raman signal enhancer.

With regard to Claim 2, Mirkin et al. teaches that the probe generates a Raman signal (paragraph 66 p. 5).

With regard to claim 5, Mirkin et al. teaches the positively charged Raman signal enhancer is a nanoparticle- Cy3-labeled alkylthiol capped oligonucleotide (Example 2, p. 10). This probe would be a composite of organic-inorganic nanoparticle (e.g. the oligonucleotide is organic and the Cy3 is inorganic).

With regard to Claims 6-7, Mirkin et al. teaches a method of detecting nucleotide occurrences at a target position wherein the position is a single nucleotide polymorphisms (abstract).

With regard to Claim 9, Mirkin et al. teaches a target segment which is equal to the combined nucleotides of the capture oligonucleotide probe and the Raman active oligonucleotide probe (Figure 4).

With regard to Claim 10, the claim is broadly interpreted to define the length of the Raman-active oligonucleotide probe as the entire length which would include the Cy3 and the nanoparticle attached to the end, Mirkin et al. teaches the probes include a A10 linker, which would make the probe longer than the target and therefore the target is less than the Raman active probe (Figure 4 and Example 4 paragraphs 162 p. 11).

With regard to Claim 13, Mirkin et al. teaches the target nucleic acid is isolated from a source and contacted to a population of capture oligonucleotide probes, but does an amplification step (Example 1 p. 10).

With regard to Claim 14, Mirkin et al. teaches a method wherein 500 pM of the nanoparticle probes are detected (e.g. less than a 1000 molecules) (p. 12 paragraph 167).

With regard to Claim 15, Mirkin et al teaches a method wherein the substrate is a biochip (p. 12 paragraph 167).

With regard to claim 16, Mirkin et al. teaches that the Raman active oligonucleotide probe is detected using SERS (abstract).

With regard to Claim 17, Mirkin et al teaches a method wherein a first population of Raman active oligonucleotide probes are contacted at a first spot and a second population of Raman active oligonucleotide probes are contacted at a second spot wherein the probe populations comprise at least one different oligonucleotide probe (Figure 7 and paragraph 14 p. 2).

With regard to Claim 37, Mirkin et al. teaches a portion of the overhang is a constituent of the target nucleic acid sequence (Figure 3A).

With regard to claim 38, Mirkin et al. teaches a nanoparticle- Cy3-labeled alkylthiol capped oligonucleotide (Example 2, p. 10). Therefore the probe comprises a tag (e.g. the alkylthiol cap).



With regard to Claims 41-42, Mirkin et al. teaches a method of aggregating nanoparticles with the Raman active probe and therefore aggregating the nanoparticles with the nucleic acid attached to the Raman signal enhancer (Figure 1).

With regard to Claim 44, Mirkin et al. teaches aggregation in the presence of a monovalent salt (p. 13 paragraph 174).

With regard to Claim 45, Mirkin et al. teaches a method wherein the nanoparticle can be silver and therefore the positive signal is carried by the Ag atom (p. 7 paragraph 116). An heteroatom is any atom that is not carbon or hydrogen and therefore the teaching in Mirkin et al. teaches all the limitations of the claim.

### **Response to Arguments**

The reply traverses the rejection of the claims. A summary of the arguments made in the reply is presented below with response to arguments following.

The reply asserts that as evidenced by the 1.132 Declaration that Mirkin et al. does not teach a positively charged Raman signal enhancer maintaining its positive charge after binding with the probe target complex (p. 13 last two paragraphs). The reply asserts that the Cy3 label loses its positive charge after bonding to the gold particle (p. 14 last paragraph).

These arguments and declaration have been fully reviewed but have not been found persuasive. The rejections have been modified but the arguments will be addressed in so far as they are relevant to the pending rejections as anticipated by Mirkin et al.

The declaration asserts that the Raman active oligonucleotide probe taught by Cao et al. is not positively charged Raman charge enhancer. The declaration asserts the binding of the Cy3 to the gold particle removes the positive charge from the Cy3 (points 3-5 p. 1-2). The declaration asserts that that as evidenced by Faulds et al. Cy3 is positively charged when bound to oligonucleotides (point 4 p. 2). The declaration asserts that the difference in Cao et al. is that there is a gold particle attached such that the Cy3 loses its charge (point 5 p. 2). The declaration provides a picture of the probe of Cao et al. and the claimed invention.

These arguments have been fully reviewed but have not been found persuasive.

The declaration asserts that the charge of Cy3 label is lost when the probe of Cao et al. is further bound to a gold nanoparticle. Applicant is referring to the positive charge of the Cy3 label, while the examiner is designating the positively charged amine group located on the Cy3 as the enhancer. The term "positively charged Raman signal enhancer" has not been defined by the instant specification or the art at the time of filing. The instant specification asserts that a positively charge enhancer can be any positively charged group that is attached to the oligonucleotides without blocking the binding of the oligonucleotide to a complementary sequence (paragraph 24 p. 6). Based upon the applicant's description any moiety which maintains a positive charge can be considered a positively charge enhancer. This would encompass the positive amine group which found on the Cy3 label.

The Cy3 dye as maintains a positive charge of the N group based upon the fact that the N group is not involved in the attachment of the Cy3 molecule either to the

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probe or to the nanoparticle. The declaration has not discuss or pointed to the structure of the Cy3 molecule to show how the positive charge of the N group is removed based upon binding to the nanoparticle.

The reply asserts that the claimed nucleic acid tag complex is not attached to a metal particle. However, the claims are not limited to nucleic acids that do not have metal particles and would encompass any nucleic acid with a positively charged Raman signal enhancer. Further the figure provided by the declaration indicates the complex is attached to Ag metal particles. The "claimed invention" figure discloses limitations which are not required by the claim, such as the Ag particles, the tag, and the positive amine group (for Claim 1). As such the figure does not have the same scope as the claimed invention and point to limitations not claimed.

### ***Claim Rejections - 35 USC § 103***

The following is a quotation of 35 U.S.C. 103(a) which forms the basis for all

obviousness rejections set forth in this Office action:

(a) A patent may not be obtained though the invention is not identically disclosed or described as set forth in section 102 of this title, if the differences between the subject matter sought to be patented and the prior art are such that the subject matter as a whole would have been obvious at the time the invention was made to a person having ordinary skill in the art to which said subject matter pertains. Patentability shall not be negated by the manner in which the invention was made.

6. This application currently names joint inventors. In considering patentability of the claims under 35 U.S.C. 103(a), the examiner presumes that the subject matter of the various claims was commonly owned at the time any inventions covered therein were made absent any evidence to the contrary. Applicant is advised of the obligation under 37 CFR 1.56 to point out the inventor and invention dates of each claim that was not commonly owned at the time a later invention was made in order for the examiner to

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consider the applicability of 35 U.S.C. 103(c) and potential 35 U.S.C. 102(e), (f) or (g) prior art under 35 U.S.C. 103(a).

7. Claim 3-4 and 8 are rejected under 35 U.S.C. 103(a) as being unpatentable over Mirkin et al. (US Patent Application Publication 2003/0211488 November 13, 2003) as evidenced by Glen Research Catalog (Catalog Number 105913 [www.glenresearch.com](http://www.glenresearch.com)) in view of Mirkin et al. (US Patent 6361944 March 26, 2002) (referred to as Mirkin B).

The teachings of Mirkin et al. are previously discussed in this office action.

However, Mirkin et al. do not teach Raman-active probes that comprise less than 5 or no purine residues.

Mirkin B teaches a method of detecting a nucleic acid using nanoparticles (Abstract). With regard to Claims 3-4, Mirkin B teaches a probe which comprises no purines (Seq Id No. 9 Figure 10).

With regard to Claim 8, Mirkin B teaches a method to detect multiple nucleotides mismatches in a target (e.g. a series of nucleotide occurrences at adjacent positions (Figure 12F).

Therefore it would have been prime facie obvious to one of ordinary skill in the art at the time the invention was made to modify the method of Mirkin et al. to include the probe with no purines as taught by Mirkin B. The ordinary artisan would have been motivated to modify the method of Mirkin et al. to include the probe with no purines as taught by Mirkin B because Mirkin B teaches that nanoparticles bearing only pyrimidine oligonucleotide bind in a sequence specific manner at purine and pyrimidine sites (Column 58 lines 15-25). Mirkin B. teaches that the binding allows for formation of

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triple-stranded complexes such that nanoparticle probes can be used for double stranded targets (Column 58 lines 15-25). Therefore the ordinary artisan would be motivated to use the probes of Mirkin B to detect double stranded targets.

Claim 11 is rejected under 35 U.S.C. 103(a) as being unpatentable over Mirkin et al. (US Patent Application Publication 2003/0211488 November 13, 2003) as evidenced by Glen Research Catalog (Catalog Number 105913 [www.glenresearch.com](http://www.glenresearch.com)) in view of Pastinen et al. (Genome Research July 2000 Vol. 10(7) p. 1031).

The teachings of Mirkin et al. are previously discussed in this office action.

However, Mirkin et al. does not teach determining the entire target nucleic acid by aligning detected target sequences.

Pastinen et al. teaches a method of genotyping by allele-specific primer extension on a microarray (abstract).

With regard to Claim 11, Pastinen et al. teaches genotyping in which using primer extension a user can determine the sequence of the extended target (Abstract). Pastinen et al. teaches using a array of a multiplex of primers each specifically near a SNP area of detections (p. 1033 1st column last sentence and second column 1st paragraph). It is obvious to the ordinary artisan to use the teaching of Pastinen et al. aligning the nucleotides detected to determine which SNPs are present on both alleles.

Therefore it would have been prime facie obvious to one of ordinary skill in the art at the time the invention was made to modify the method of Mirkin et al. to include the step of sequencing the target as taught by Pastinen et al. The ordinary artisan would have been motivated to modify the method of Mirkin et al. to include the step of sequencing the target as taught by Pastinen et al. a method to perform high-throughput genotyping of samples in a parallel analysis method. The ordinary artisan would be motivated to use the method of Pastinen et al. to sequence the entire target in a quick assay to determine the entire sequence of the target.

Claim 12 is rejected under 35 U.S.C. 103(a) as being unpatentable over Mirkin et al. (US Patent Application Publication 2003/0211488 November 13, 2003) as evidenced by Glen Research Catalog (Catalog Number 105913 [www.glenresearch.com](http://www.glenresearch.com)) in view of Lane et al. (US Patent 5,770,365 June 23, 1998).

The teachings of Mirkin et al. are previously discussed in this office action.

However, Mirkin et al. does not teach ligating the capture oligonucleotide probes to Raman-active oligonucleotide probes that bind to an adjacent segment of the target nucleic acid.

Lane et al. teaches a method of using nucleic acid capture moieties to detect nucleic acid sequences (Column 4, lines 19-25). Lane et al. teaches a labeled probe complementary to a target-complementary region of the capture moiety that is immobilized on insoluble support (Column 11, lines 30-35). With regard to Claim 12,

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Lane et al. teaches a method in which the detectable probe is ligated to the capture probe (a duplex-binding ligand binding site) (Figure 3).

Therefore it would have been prime facie obvious to one of ordinary skill in the art at the time the invention was made to modify the method of Mirkin et al. to further include the use ligated probes as taught by Lane et al. The ordinary artisan would have been motivated to improve the method of Mirkin et al. because Lane et al. teaches that the ligation method can be used for the detection of nucleic acid sequences, which do not occur near the terminus of an intact target strand (Column 12, lines 15-20).

8. Claims 22-24, 26-27, 29-32 are rejected under 35 U.S.C. 103(a) as being unpatentable over Mirkin et al. (US Patent Application Publication 2003/0211488 November 13, 2003) as evidenced by Glen Research Catalog (Catalog Number 105913 [www.glenresearch.com](http://www.glenresearch.com)) in view of Chan et al. (US Patent Application Publication March 27, 2003) and Corbierre et al. (Journal of American Chem. Soc 2001 Vol. 123 p. 10411).

With regard to Claim 22, Mirkin et al. teaches a multiplexed detection method of oligonucleotide targets bound to capture probes and detected using SERS (surface enhanced Raman) Raman probes (Abstract, p. 3 paragraphs 45-49). Mirkin et al. teaches contacting a target nucleic acid with a plurality of capture probes bound to a substrate forming a single-stranded overhang (Abstract, p. 3 paragraphs 45-49). Mirkin et al. teaches contacting the probe-target with a population of Raman-active oligonucleotides which forms a three-component sandwich assay used in a microarray

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(e.g. biochip) format composed of nanoparticle probes (Raman probes) detecting a bound target: capture probe duplex (Abstract, p. 3 paragraphs 45-49 ).

Mirkin et al. teaches a nanoparticle- Cy3-labeled alkylthiol capped oligonucleotide (Example 2, p. 10). The positive amine group on the Cy3 label would be considered a positively charged enhancer.

With regard to Claim 23, Mirkin et al. teaches that a florescent signal is detected (Figure 8).

With regard to Claim 26, Cao et al. teaches that a Raman spectra is detected (abstract).

With regard to Claim 27, Cao et al. teaches comparing the signal to standard known Raman spectra labels (Figure 8). Therefore Cao et al. compares the detected spectra with known spectrum to identify the nucleotide occurrence.

However, Mirkin et al. do not teach a method of labeling the target with two labels, applying premade aggregates of metallic colloids to the probe-target, and applying an alternating current.

Chan et al. teaches a method for spatial resolution of signal detection (Abstract). With regard to Claim 22, Chan et al. teaches a method of passing a target through an optical detector to read florescent signals (p. 12 paragraphs 114 and 115). Chan et al. teaches the probe can be labeled with FRET labels (e.g. two labels on the probe) (paragraph 148 p. 16). Chan et al. teaches that the target nucleotide is pulled through the nanoslit of the channel by applying an alternating current (AC current) filed to the metal layer (p. 14 paragraph 132).



With regard to Claim 24, Chan et al. teaches the probe can be labeled with FRET labels (paragraph 148 p. 16).

With regard to Claim 29, Chan et al. teaches determining a series of nucleotide occurrences for one target by determination of a population of labeled probes (Figure 2 and paragraph 41 p. 4).

With regard to Claim 30, Chan et al. teaches passing the complexes through an optical detector to read the fluorescent signal (p. 12 paragraph 115).

With regard to Claim 31, Chan et al. teaches an interactor station comprised of the channel and the optical detector (e.g. a microelectromechanical system) (p. 12 paragraph 115).

With regard to Claim 32, Chan et al. teaches that the target nucleotide is pulled through the nanoslit of the channel by applying an alternating current (AC current) filed to the metal layer (p. 14 paragraph 132). Chan et al. teaches the optical system uses radiation modulated frequencies (AC current oscillations) in the range of 10 MHz to 1 GHz (p. 15 paragraph 138).

With regard to Claim 22, Corbierre et al. teaches a method of synthesizing nanoparticles such as gold before incorporation (p. 10411 2<sup>nd</sup> paragraph). Corbierre et al. teaches a method of making pre-made nanoparticles (p. 10411 2<sup>nd</sup> paragraph).

Therefore it would have been prime facie obvious to one of ordinary skill in the art at the time the invention was made to modify the method of Cao et al. and Mirkin et al. to further include the use of a AC current and two label FRET system as taught by Chan et al. and premade gold nanoparticles as taught by Corbierre et al. The ordinary

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artisan would have been motivated to modify the method of Cao et al. and Mirkin et al. to further include the use of a AC current and two label FRET system as taught by Chan et al. because Chan et al. teaches a method of linear analysis of DNA which can allow for the development of specific sequences to be used in sequence-specific tagging and differentially tagging to increase resolution (p. 1 paragraph 3 and 4). The ordinary artisan would have been motivated to modify the method of Cao et al. and Mirkin et al. to further include the use of a premade gold nanoparticles as taught by Corbierre et al., because Corbierre et al. teaches that premade nanoparticles provides full synthetic control over the making of the nanoparticle (p. 10412 last paragraph).

9. Claim 25 is rejected under 35 U.S.C. 103(a) as being unpatentable over Mirkin et al. (US Patent Application Publication 2003/0211488 November 13, 2003) as evidenced by Glen Research Catalog (Catalog Number 105913 [www.glenresearch.com](http://www.glenresearch.com)) in view of Chan et al. (US Patent Application Publication March 27, 2003) and Corbierre et al. (Journal of American Chem. Soc 2001 Vol. 123 p. 10411) as applied to claims 22-24, 26-27, and 29-32 above and further in view of Bruchez, Jr. et al. (US Patent Application 09/815585 March 21, 2002).

Neither Mirkin et al. or Chan et al. or Corbierre et al. teach FRET labels of TAMRA and ROX.

With regard to Claim 25, Bruchez, Jr. et al. teaches that the fluorophores, which can be used as labels, include TAMRA and ROX (p. 13 paragraph 151).

Therefore it would have been prime facie obvious to one of ordinary skill in the art at the time the invention was made to modify the method of Mirkin et al. , Chan et al., and Corbierre et al. to further include any type of FRET labels including TAMRA and ROX as presented by Bruchez Jr. et al. The use of FRET labels is well known in the art and the use of different types of FRET labels are interchangeable. Therefore the ordinary artisan would use any type of FRET label for the method of Mirkin et al., Chan et al., and Corbierre et al. including TAMRA and ROX to detect nucleotide occurrences on a target strand.

10. Claim 28 is rejected under 35 U.S.C. 103(a) as being unpatentable over Mirkin et al. (US Patent Application Publication 2003/0211488 November 13, 2003) as evidenced by Glen Research Catalog (Catalog Number 105913 [www.glenresearch.com](http://www.glenresearch.com)) in view of Chan et al. (US Patent Application Publication March 27, 2003) and Corbierre et al. (Journal of American Chem. Soc 2001 Vol. 123 p. 10411) as applied to claims 22-24, 26-27, and 29-32 above and further in view of Livak et al (US Patent 5723591 March 3, 1998)

Neither Mirkin et al. or Chan et al. or Corbierre et al. teach the two labels are located about 3-6 nm apart.

With regard to Claim 28, Livak et al. teaches that the quencher molecule and reporter should be between 6-16 nucleotides (Column 3, line 63). The distance between nucleotides is 0.23 nm, therefore the distance between a reporter and quencher can be between 1.38 to 3.68 nm apart (between 3-6 nm).

Therefore it would have been prime facie obvious to one of ordinary skill in the art at the time the invention was made to modify the method of Mirkin et al., Chan et al., and Corbierre et al. to further include distance limitation as taught by Livak et al. The ordinary artisan would have been motivated to modify the method of Mirkin et al., Chan et al., and Corbierre et al. to further include distance limitation as taught by Livak et al. because Livak et al. teaches that there is a distance that must be maintained between the quencher and reporter in order for the quencher to be able to quench the reporter in the assay (Column 3, lines 60-65).

11. Claims 33-34 and 43 are rejected under 35 U.S.C. 103(a) as being unpatentable over Mirkin et al. (US Patent Application Publication 2003/0211488 November 13, 2003) as evidenced by Glen Research Catalog (Catalog Number 105913 [www.glenresearch.com](http://www.glenresearch.com)) in view of Alivisatos et al. (US Patent 6884478 April 26, 2005).

Mirkin et al. teaches a multiplexed detection method of oligonucleotide targets bound to capture probes and detected using SERS (surface enhanced Raman) Raman probes (Abstract, p. 3 paragraphs 45-49). Mirkin et al. teaches a method in which the probe is a nanoparticle- Cy3-labeled alkylthiol capped oligonucleotide (Example 2, p. 10). The amine group of the Cy3 label would be considered a positively charged enhancer.

Further Mirkin et al. teaches that nanoparticles can be made of silver or TiO<sub>2</sub> (p. 7 paragraph 116) which are both positively charged. The instant specification asserts

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that a positively charge enhancer can be any positively charged group that is attached to the oligonucleotides without blocking the binding of the oligonucleotide to a complementary sequence (paragraph 24 p. 6). The nanoparticle does not block the binding of the probe and the complementary sequence, has a positive charge and therefore is also a positively charged enhancer.

Mirkin et al. teaches irradiating the nucleic acid with light and detecting the Raman signal (Abstract, p. 3 paragraphs 45-49).

With regard to Claim 34, Mirkin et al. teaches that the amine group of the Cy3 is charged (as evidenced by Glen Research Catalog).

However, Mirkin et al. do not teach Raman-active probes that comprise primary amines with alkyl chain of 1 to 25 carbon atoms, but rather alkylthiols.

With regard to Claims 33 and 43, Alivisatos et al. teaches that nanoparticles can be treated with alkylamines, alkylthiols, or carboxylic acid functional groups to make them soluble in water (column 5, lines 25-32)

Therefore it would have been prime facie obvious to one of ordinary skill in the art at the time the invention was made to modify the method of Mirkin et al. to include nanoparticles with alkylamines as taught by Alivisatos et al. It would have been obvious to one of ordinary skill in the art at the time the invention was made to choose from a finite number of predictable alkyl functionalized groups to coat the nanoparticle including alkylamines with a reasonable expectation of success of producing a functionalized nanoparticle which can be soluble in water.

12. Claims 36 is rejected under 35 U.S.C. 103(a) as being unpatentable over Mirkin et al. (US Patent Application Publication 2003/0211488 November 13, 2003) as evidenced by Glen Research Catalog (Catalog Number 105913 [www.glenresearch.com](http://www.glenresearch.com)) in view of Alivisatos et al. (US Patent 6884478 April 26, 2005) as applied to Claims 33-34 and in view of Mirkin et al. (US Patent 6361944 March 26, 2002) (referred to as Mirkin B).

However, Mirkin et al. and Alivisatos et al. do not teach Raman-active probes that comprise no purine residues.

Mirkin B teaches a method of detecting a nucleic acid using nanoparticles (Abstract). With regard to Claims 3-4, Mirkin B teaches a probe which comprises no purines (Seq Id No. 9 Figure 10).

With regard to Claim 36, Mirkin B teaches a method to detect multiple nucleotides mismatches in a target (e.g. a series of nucleotide occurrences at adjacent positions (Figure 12F).

Therefore it would have been prime facie obvious to one of ordinary skill in the art at the time the invention was made to modify the method of Mirkin et al. to include the probe with no purines as taught by Mirkin B. The ordinary artisan would have been motivated to modify the method of Mirkin et al. to include the probe with no purines as taught by Mirkin B because Mirkin B teaches that nanoparticles bearing only pyrimidine oligonucleotide bind in a sequence specific manner at purine and pyrimidine sites (Column 58 lines 15-25). Mirkin B. teaches that the binding allows for formation of triple-stranded complexes such that nanoparticle probes can be used for double

stranded targets (Column 58 lines 15-25). Therefore the ordinary artisan would be motivated to use the probes of Mirkin B to detect double stranded targets.

### ***Double Patenting***

**13.** The nonstatutory double patenting rejection is based on a judicially created doctrine grounded in public policy (a policy reflected in the statute) so as to prevent the unjustified or improper timewise extension of the “right to exclude” granted by a patent and to prevent possible harassment by multiple assignees. A nonstatutory obviousness-type double patenting rejection is appropriate where the conflicting claims are not identical, but at least one examined application claim is not patentably distinct from the reference claim(s) because the examined application claim is either anticipated by, or would have been obvious over, the reference claim(s). See, e.g., *In re Berg*, 140 F.3d 1428, 46 USPQ2d 1226 (Fed. Cir. 1998); *In re Goodman*, 11 F.3d 1046, 29 USPQ2d 2010 (Fed. Cir. 1993); *In re Longi*, 759 F.2d 887, 225 USPQ 645 (Fed. Cir. 1985); *In re Van Ornum*, 686 F.2d 937, 214 USPQ 761 (CCPA 1982); *In re Vogel*, 422 F.2d 438, 164 USPQ 619 (CCPA 1970); and *In re Thorington*, 418 F.2d 528, 163 USPQ 644 (CCPA 1969).

A timely filed terminal disclaimer in compliance with 37 CFR 1.321(c) or 1.321(d) may be used to overcome an actual or provisional rejection based on a nonstatutory double patenting ground provided the conflicting application or patent either is shown to be commonly owned with this application, or claims an invention made as a result of activities undertaken within the scope of a joint research agreement.

Effective January 1, 1994, a registered attorney or agent of record may sign a terminal disclaimer. A terminal disclaimer signed by the assignee must fully comply with 37 CFR 3.73(b).

14. Claims 1-17, 22-34, 36-38, 41-45 are provisionally rejected on the ground of nonstatutory obviousness-type double patenting as being unpatentable over claim 1-4 of copending Application No. 11/414,611. Although the conflicting claims are not identical, they are not patentably distinct from each other. Claims 1, 4, 33, and 43 of the pending application are drawn to a method comprising a light source, a nucleic acid comprising a positively charged enhancer which is an amine, and detection of the Raman signal, which are identical in steps to Claims 1-4 of application 11/414,611.

This is a provisional obviousness-type double patenting rejection because the conflicting claims have not in fact been patented.

#### **Response to arguments**

The reply requests the double patent rejections be held in abeyance until the indication of allowable subject matter. As such the double patenting rejections have been maintained.

#### **Conclusion**

15. No Claims are allowed.

16. Any inquiry concerning this communication or earlier communications from the examiner should be directed to Katherine Salmon whose telephone number is (571) 272-3316. The examiner can normally be reached on Monday-Friday 8AM-430PM.

If attempts to reach the examiner by telephone are unsuccessful, the examiner's supervisor, Ram Shukla can be reached on (571) 272-0735. The fax phone number for the organization where this application or proceeding is assigned is 571-273-8300.



Art Unit: 1634

Information regarding the status of an application may be obtained from the Patent Application Information Retrieval (PAIR) system. Status information for published applications may be obtained from either Private PAIR or Public PAIR. Status information for unpublished applications is available through Private PAIR only. For more information about the PAIR system, see <http://pair-direct.uspto.gov>. Should you have questions on access to the Private PAIR system, contact the Electronic Business Center (EBC) at 866-217-9197 (toll-free). If you would like assistance from a USPTO Customer Service Representative or access to the automated information system, call 800-786-9199 (IN USA OR CANADA) or 571-272-1000.

/Katherine Salmon/  
Examiner, Art Unit 1634

/Juliet C Switzer/

Primary Examiner, Art Unit 1634